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## Determination of partition coefficients of non-ionic contrast agents by reversed-phase high-performance liquid chromatography

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### ABSTRACT

Reversed-phase high-performance liquid chromatography was used to measure the lipophilicities of non-ionic contrast agents. Calculated partition coefficients were correlated with the capacity factors extrapolated to zero organic modifier content.

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### INTRODUCTION

Studies of quantitative structure–activity and structure–toxicity relationships<sup>1,2</sup> have shown that the octanol–water partition coefficient ( $P$ ) is one of the most important physical parameters related to the biological activities and toxicities of organic compounds. The shake-flask method<sup>3,4</sup> is usually used for the determination of  $\log P$ . However, this method is tedious and not simple. Reversed-phase high-performance liquid chromatography (RP-HPLC) has also been used to determine  $\log P$ , as it is simple, rapid and accurate. Octadecylsilica with<sup>5,6</sup> or without<sup>7–11</sup> previous treatment with trimethylsilyl chloride is the most widely used stationary phase.

The method involves:

(a) a linear correlation between capacity factor ( $\log k'$ ) and organic modifier volume fraction ( $\varphi$ ):

$$\log k' = \log k'_w + S\varphi \quad (1)$$

where

$$\varphi = V_{\text{Org. modifier}} / (V_{\text{Org. modifier}} + V_{\text{water}}) \quad (2)$$

the intercept ( $\log k'_w$ ) is the capacity factor extrapolated to zero organic modifier content and the slope,  $S$ , is the slope parameter<sup>12</sup>;

(b) a linear regression between  $\log k'_w$  and  $\log P$  for several compounds (training

set) with known partition coefficients, usually determined by the shake-flask method:

$$\log P = a + b \log k'_w \quad (3)$$

(c) the determination of the  $\log k'$  and  $\log k'_w$  values for the test compounds by chromatography; the  $\log P$  values of the test compounds are obtained from eqn. 3.

In eqn. 3, partition conditions are represented by a value of  $b$  close to unity. However, a search for a chromatographic system giving a regression line in which a large change in  $\log P$  corresponds to a small modification of  $\log k'$  ( $b > 1$ ) is especially important. A value of  $b$  of about 2 may be useful<sup>1</sup>.

On the other hand, calculation methods<sup>13-17</sup> could be used to avoid the experimental determination of  $\log P$ .

In connection with our work on non-ionic contrast agents, several 5-amino-2,4,6-triiodoisophthalic and 3,5-diamino-2,4,6-triiodobenzoic acid derivatives have been prepared. As part of a study of the physico-chemical properties of the contrast agents, we report here the determination of their chromatographic parameters  $\log k'_w$  and  $S$ , and the relationship between  $\log k'_w$  and calculated partition coefficients.

## EXPERIMENTAL

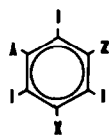
### Materials

HPLC-grade acetonitrile were obtained from Fluka (Buchs, Switzerland). Contrast agents 3-7 (Table I) were prepared by us<sup>18,19</sup>. Compounds 1 (iohexol) and 2 (iopamidol) were isolated from commercially available pharmaceutical products.

### Chromatography

The HPLC instrument consisted of a Hewlett-Packard (Waldbronn Analytical Division, Waldbronn, F.R.G.) chromatograph with an autosampler and an

TABLE I  
TRAINING SET OF CONTRAST AGENTS



Compound	X	Y <sup>a</sup>	Z <sup>a</sup>
1	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH
2	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	NHCOCH(OH)CH <sub>3</sub> (L) <sup>b</sup>
3	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH
4	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>3</sub>
5	CONHC(CH <sub>2</sub> OH) <sub>3</sub>	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>3</sub>
6	CONHCH <sub>2</sub> CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH
7	CON(CH <sub>3</sub> )CH <sub>2</sub> CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH

<sup>a</sup> Ac = Acetyl.

<sup>b</sup> (L) = Chiral center configuration of the lactoyl group.

HP 1090 A detection system, operating at 254 nm, HP 85 B computer, HP 9121 disc drive and HP Thinkjet printer.

A reversed-phase Novapack C<sub>18</sub> column (15 cm × 3.7 mm I.D., 4 μm particle size) (Millipore-Waters, Milford, MA, U.S.A.) was used.

Retention times were measured by injecting 5 μl of an aqueous contrast agent solution (1 mg/ml) and eluting under isocratic conditions with several acetonitrile-water volume fractions (eqn. 2). The column temperature was 40°C in order to ensure adequate thermostating and good reproducibility of the chromatographic data. Two flow-rates, 0.5 and 1.0 ml/min, were used. The column dead time,  $t_0$ , was determined at each flow-rate and  $\phi$  used by injecting 3% sodium nitrate solution as the non-retained compound. The capacity factor,  $k'$ , is defined as

$$k' = (t_R - t_0)/t_0 \quad (4)$$

where  $t_R$  is the mean and weighted retention time of the test compound.

The experimental conditions were chosen in order to obtain short retention times ( $t_R$ ) without losing the discrimination power between the different contrast agents. Hence broad chromatography peaks and thus inaccurate determinations of  $t_R$  can be avoided. Fortunately, owing to the high aqueous solubility of the contrast agents, it was possible to work with small  $t_R$  and  $\phi$  values in order to obtain a linear correlation according to eqn. 1.

## RESULTS AND DISCUSSION

### Log $P$ calculation

The Hansch-Leo method<sup>14</sup> is the most generally used procedure for the calculation of log  $P$  values. They suggested a group contribution method based on fragment  $f_i$  and corrective factor  $F_j$  values:

$$\log P = \sum_i a_i f_i + \sum_j b_j F_j \quad (5)$$

For molecules as complex as the contrast agents, with numerous inter- and intramolecular interactions, the calculated log  $P$  values deviated from the experimental results. This was evident when we used the experimental log  $P$  values obtained from Haavaldsen *et al.*<sup>20</sup> to evaluate the fit of this parameter using the Hansch-Leo procedure<sup>14</sup> (Table II), the log  $P$  values calculated by the Hansch-Leo method being much more positive than the experimental values. This does not mean that this method is invalid but implies that as we can not modify the group contributions  $f_i$  in eqn. 5, the  $F$  correction factors, especially the proximity factors ( $F_p$ ), are overestimated and must be corrected.

The modifications made in this work in order to obtain a good correlation between the calculated and experimental values were as follows: (1)  $F_{p3}$  proximity factors were not considered, except for X and/or Y = CONHC(CH<sub>2</sub>OH)<sub>3</sub>; (2) all  $F_{p2}$  factors of the amido and carbamoyl groups with a hydroxyl moiety were considered, except for X and/or Y = CONHCH(CH<sub>2</sub>OH)<sub>2</sub>, where this contribution was divided

TABLE II  
CALCULATED AND REPORTED LOG *P* VALUES

Compound	X	Y	Z
1	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH
8	CON(CH <sub>3</sub> )CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CON(CH <sub>3</sub> )CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	NHCOCH <sub>3</sub>
9	CON(CH <sub>3</sub> )CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CON(CH <sub>3</sub> )CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH <sub>2</sub> OH
10	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	NHCOCH <sub>3</sub>
11	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH <sub>2</sub> OH
12	CONHCH <sub>2</sub> CH <sub>2</sub> OH	CONHCH <sub>2</sub> CH <sub>2</sub> OH	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH
13	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	CONHCH <sub>2</sub> CH(OH)CH <sub>2</sub> OH	N(Ac)CH <sub>3</sub>
14	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	N(Ac)CH <sub>2</sub> CH <sub>2</sub> OH
15	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	CONHCH(CH <sub>2</sub> OH) <sub>2</sub>	N(Ac)CH <sub>2</sub> CH(OH)CH <sub>2</sub> OH

<sup>a</sup> From ref. 20.

<sup>b</sup> Calculated according to the Hansch-Leo method.

<sup>c</sup>  $\Delta^* = \log P_{\text{obs}} - \log P_{\text{HL}}$ ;  $\Delta^{**} = \log P_{\text{obs}} - \log P_{\text{CHL}}$ ;  $\Delta^{***} = \log P_{\text{obs}} - \log P_{\text{calc}}$ .

<sup>d</sup> Calculated according to the corrected Hansch-Leo method.

<sup>e</sup> From eqn. 7.

by a factor of two; and (3) the hydroxyl-hydroxyl  $F_{p_2}$  factors were calculated from the following empirical equation, obtained by a trial and error procedure:

$$F'_{p_2}(\text{OH}, \text{OH}) = [(A - B)C/N]F_{p_2}(\text{OH}, \text{OH}) \quad (6)$$

where *A* is the number of chains with two or more OH groups, *B* is the number of chains with less than two OH groups, *C* is the number of chains with OH and *N* is the total number of hydroxyl groups.

We modified the contribution of the  $F_{p_2}(\text{OH}, \text{OH})$  factors to consider both the number and the molecular distribution of the OH groups. Hence there are two factors in eqn. 6: (a) *C/N* can unmodify or decrease the magnitude of the  $F_{p_2}(\text{OH}, \text{OH})$ ; and (b) (*A - B*) can unmodify, increase or decrease the magnitude of the  $F_{p_2}(\text{OH}, \text{OH})$  and even reverse the sign of this contribution (*A < B*), which is always positive in the Hansch-Leo method.

The following equation shows a good correlation between log *P* calculated as above and reported experimental values<sup>20</sup>:

$$\log P_{\text{obs}} = 0.067 + 1.053 \log P_{\text{CHL}} \quad (7)$$

$$n = 9; r = 0.966; SEE = 0.101; F(1,7) = 97.82; p < 0.0001.$$

where *SEE* = standard error of estimation; *n* = number of data points (compounds); *r* = correlation coefficient; *F* = *F*-statistic significance test with 1 and 7 degrees of freedom; *p* = observed significance level of *F* (probability).

#### Log *P* determination

Table III gives the capacity factors at different organic modifier volume fractions (log  $k'_o$ ) obtained with flow-rates of 0.5 and 1 ml/min. In the latter instance,

$\text{Log } P_{\text{obs}}^a$	$\text{Log } P_{\text{HL}}^b$	$\Delta^{**c}$	$\text{Log } P_{\text{CHL}}^d$	$\Delta^{***c}$	$\text{Log } P_{\text{calc}}^e$	$\Delta^{****c}$
-3.05	-1.71	-1.34	-2.99	-0.06	-3.08	0.03
-2.17	-0.82	-1.35	-2.10	-0.07	-2.14	-0.03
-2.28	-1.18	-1.10	-2.28	0.00	-2.33	0.05
-2.27	0.10	-2.37	-2.17	-0.10	-2.22	-0.05
-2.47	-1.24	-1.23	-2.34	-0.13	-2.40	-0.07
-1.86	-0.32	-1.54	-1.81	-0.05	-1.84	-0.02
-2.05	-0.81	-1.24	-2.09	0.04	-2.13	0.08
-2.33	-0.25	-2.08	-2.43	0.10	-2.49	0.16
-2.80	-0.73	-2.07	-2.57	-0.23	-2.64	-0.16

the acetonitrile concentration can be decreased to 5% ( $\phi = 0.05$ ) without increasing the retention times too much.

Fig. 1 shows the linear correlations of  $\phi$  with  $\log k'$  obtained at a flow-rate of 0.5 ml/min for compounds 1-7. Table IV gives the linear regression data for the correlations and also the  $\log P$  values calculated by the corrected Hansch-Leo method. The intercept  $\log k'_w$  shows the degree of affinity of the compound for the lipophilic phase when aqueous elution occurs. The slope  $S$  shows the reduction in the affinity of the compound for the stationary phase with increase in the organic modifier concentration.

The relationship between  $\log k'_w$  and calculated partition coefficients,  $\log P_{\text{CHL}}$ , is expressed by the following equations:

$$\log P_{\text{CHL}} = -2.113 + 1.813 \log k'_w \quad (8)$$

$$n = 7; r = 0.980; SEE = 0.221; F(1,5) = 123.38; p < 0.001$$

and

$$\log P_{\text{CHL}} = -2.244 + 2.007 \log k'_w \quad (9)$$

$$n = 6; r = 0.998; SEE = 0.072; F(1,4) = 1098.55; p < 0.001.$$

The data referring to these equations are given in Table IV. Eqn. 9 is obtained from the same data as eqn. 8, excluding the most deviating point (residual = 0.384) corresponding to iopamidol. It is noteworthy that there is an improvement in the quality of the regression on going from eqn. 8 to 9.

The "deviant" behaviour of iopamidol could be explained by its structural dissimilarities with the other compounds in the training set. Moreover, the calculated  $\log k'_w$  values for iohexol (-0.367) and iopamidol (-0.366) were almost identical and

TABLE III  
 RP-HPLC CAPACITY FACTORS ( $\text{LOG } k'_p$ ) OF CONTRAST AGENTS

Compound	Flow-rate 0.5 ml/min					Flow-rate 1.0 ml/min				
	$\text{Log } k'_{0.15}$	$\text{Log } k'_{0.20}$	$\text{Log } k'_{0.25}$	$\text{Log } k'_{0.30}$	$\text{Log } k'_{0.05}$	$\text{Log } k'_{0.10}$	$\text{Log } k'_{0.15}$	$\text{Log } k'_{0.20}$	$\text{Log } k'_{0.25}$	
1	-0.887	-1.100	-1.196	-1.439	-1.221	-1.561	-1.309	-1.201	-1.291	
2	-0.917	-1.194	-1.303	-1.524	-0.983	-1.505	-1.353	-1.318	-1.439	
3	-0.600	-0.906	-1.009	-1.243	-0.373	-1.196	-1.077	-1.055	-1.047	
4	-0.164	-0.509	-0.732	-0.910	0.058	-0.596	-0.484	-0.613	-0.732	
5	0.384	0.042	-0.242	-0.460	0.699	0.015	0.096	-0.016	-0.250	
6	-0.478	-0.788	-1.044	-1.336	-0.437	-0.974	-0.814	-0.838	-1.064	
7	-0.383	-0.647	-0.861	-1.149	-0.166	-0.796	-0.729	-0.824	-0.884	

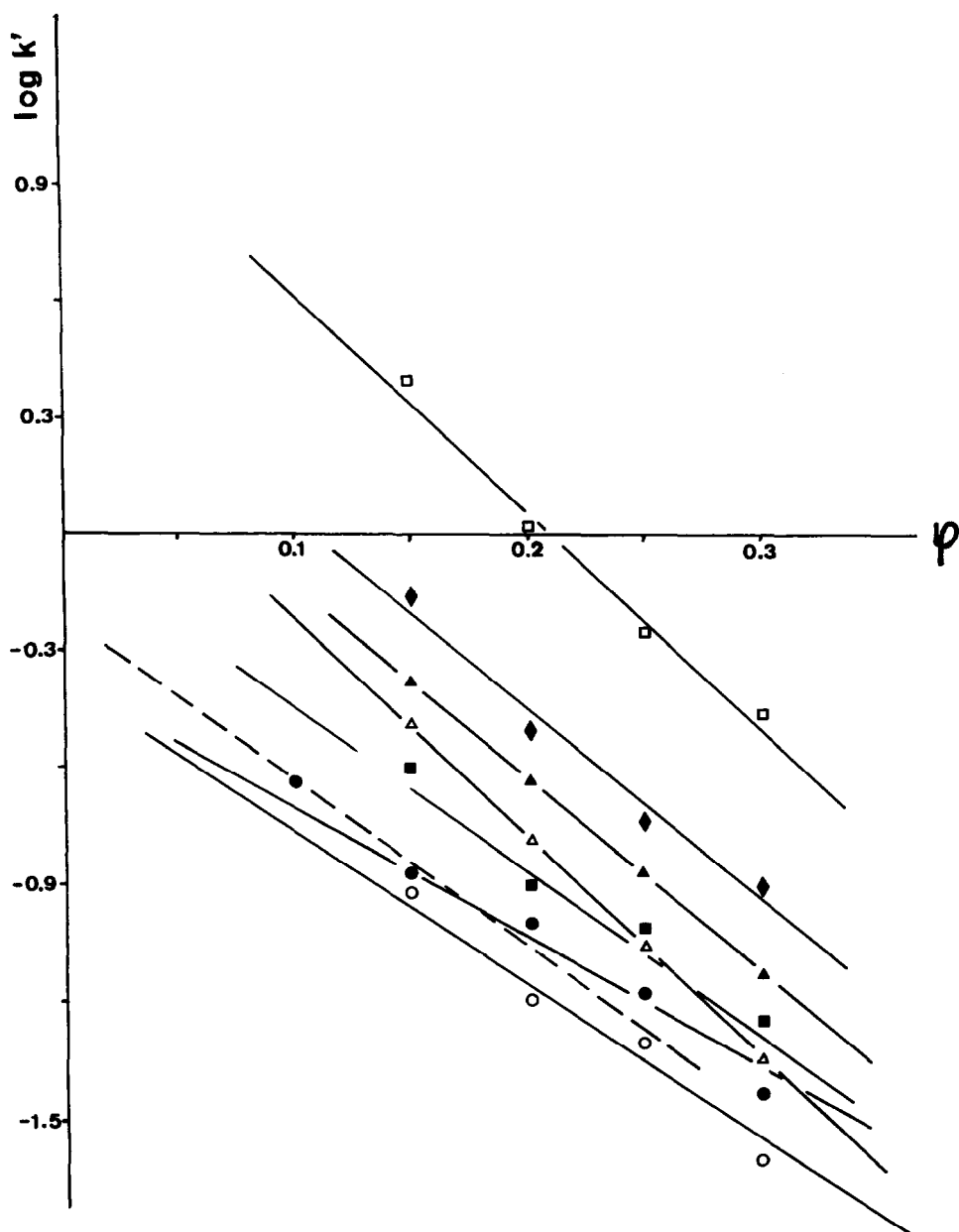


Fig. 1. Relationship between  $\log k'$  values of CA and acetone nitrile concentration ( $\phi$ ) in the mobile phase (flow-rate 0.5 ml/min). The compounds are numbered as in Table I. Key:  $\circ$  = 1;  $\bullet$  = 2;  $\blacksquare$  = 3;  $\blacklozenge$  = 4;  $\square$  = 5;  $\triangle$  = 6;  $\blacktriangle$  = 7.

hence the partition coefficients calculated by eqn. 8 and 9 were also the same. This result conflicts with the experimental data found by Haavaldsen *et al.*<sup>20</sup> for iohexol ( $\log P = -3.046$ ) and by Jacobsen<sup>21</sup> for the iohexol ( $\log P = -3.000$ ) and iopamidol

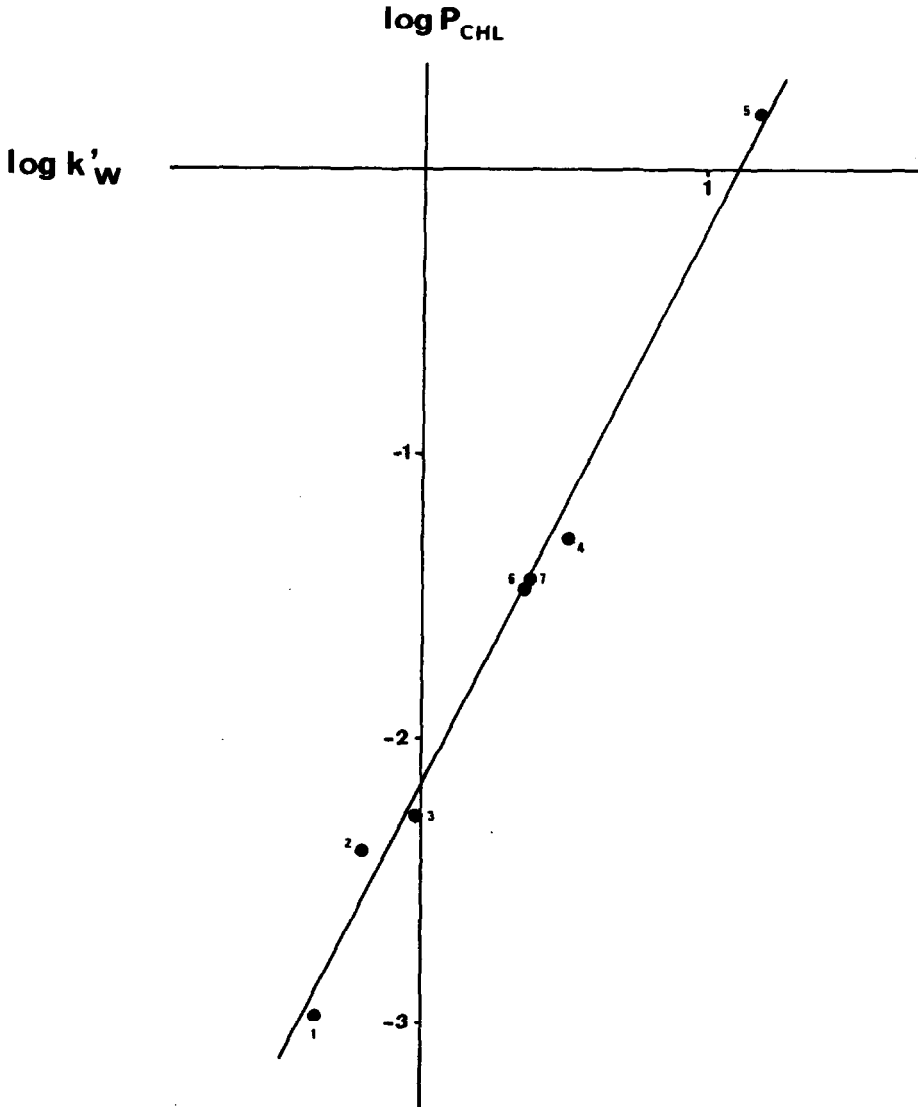


Fig. 2. Correlation of calculated  $\log P_{\text{CHL}}$  with  $\log k'_w$ . Regression data are given in eqn. 10.

( $\log P = -2.699$ ). To obviate this discrepancy,  $\log k'_w$  of iopamidol was recalculated using three data points at  $\varphi = 0.10, 0.15$  and  $0.20$ , giving  $\log k'_w = -0.208$ . By including this new  $\log k'_w$  value, the following equation is obtained (Fig. 2):

$$\log P_{\text{CHL}} = -2.185 + 1.931 \log k'_w \quad (10)$$

$$n = 7; r = 0.994; \text{SEE} = 0.120; F(1,5) = 428.67; p < 0.001.$$



TABLE IV

LOG  $P_{\text{CHL}}$  VALUES OF THE TRAINING SET AND LINEAR REGRESSION DATA FOR THE CORRELATIONS OF LOG  $k'$  AND  $\varphi$ 

Compound	Log $P_{\text{CHL}}$	Calculated for $\log k' = \log k'_w + S\varphi$					
		Log $k'_w$	$S$	$r$	$SEE^a$	$F(1,2)$	$n$
1	-2.988 <sup>b</sup>	-0.367	-3.504	-0.988	0.043	82.74 <sup>d</sup>	4
2	-2.393 <sup>c</sup>	-0.366	-3.860	-0.988	0.048	79.19 <sup>d</sup>	4
3	-2.267	-0.025	-4.064	-0.984	0.058	60.07 <sup>d</sup>	4
4	-1.308	0.529	-4.922	-0.988	0.060	83.32 <sup>d</sup>	4
5	0.191	1.198	-5.632	-0.995	0.044	206.12 <sup>d</sup>	4
6	-1.484	0.362	-5.660	-0.999	0.016	1647.92 <sup>e</sup>	4
7	-1.457	0.370	-5.024	-0.999	0.021	691.30 <sup>e</sup>	4

<sup>a</sup>  $SEE$  = standard error of estimate.<sup>b</sup> Reported values: -3.046<sup>20</sup>; -3.000<sup>21</sup>.<sup>c</sup> Reported value: -2.699<sup>21</sup>.<sup>d</sup>  $p \leq 0.01$ .<sup>e</sup>  $p \leq 0.001$ .

Log  $P$  values of compounds 1-7 obtained using eqn. 10 are -2.894 (1), -2.587 (2), -2.233 (3), -1.164 (4), +0.128 (5), -1.486 (6) and -1.471 (7). As can be seen, log  $P$  for iohexol and iopamidol calculated by the corrected Hansch-Leo method (Table IV) and by eqn. 10 are close to the literature reported values (Table IV).

On the other hand, as can be seen in Table III, the log  $k'$  values obtained at a flow-rate of 1 ml/min were poorly correlated with  $\varphi$ . This shows that with highly water-soluble compounds such as the contrast agents, an increase in flow-rate may cause a deviation of the partition phenomenon, probably owing to the poor retention of the compounds under these conditions.

## CONCLUSION

The measurement of partition coefficients of highly water-soluble contrast agents by RP-HPLC is a viable alternative to the tedious shake-flask method. It is possible to calculate log  $P$  values of molecules as complex as the contrast agents by making slight modifications to the Hansch-Leo method. The calculated log  $P$  values showed a high correlation with experimental log  $P$  values (eqn. 7) and with log  $k'_w$  (eqns. 9 and 10). As Leo reported<sup>22</sup>, the deviation between calculated log  $P$  and values determined by RP-HPLC may be due more to unsuitable experimental conditions than to a formal error in the calculation procedure.

Once it has been verified that the method used for the calculation of log  $P$  affords accurate results for a set of compounds, the use of calculated log  $P$  values correlated with log  $k'_w$  values for the training set allows experimental log  $P$  values for compounds not included in that set to be obtained.

## REFERENCES

- 1 H. Terada, *Quant. Struct.-Act. Relat.*, 5 (1986) 81.
- 2 R. Franke, *Theoretical Drug Design Methods*, Elsevier, Amsterdam, 1984.
- 3 J. J. Kirchner, W. E. Acree, A. J. Leo and G. Gelli, *J. Pharm. Sci.*, 74 (1985) 1129.
- 4 M. Kuchar, E. Kraus, M. Jelinková, V. Rejhlee and V. Miller, *J. Chromatogr.*, 347 (1985) 335.
- 5 M. S. Mirrlees, S. J. Moulton, C. T. Murphy and P. J. Taylor, *J. Med. Chem.*, 19 (1976) 615.
- 6 J. M. McCall, *J. Med. Chem.*, 18 (1975) 649.
- 7 T. L. Hafkenschied and E. Tomlinson, *Adv. Chromatogr.*, 25 (1986) 1.
- 8 M. Recatanini, *Quant. Struct.-Act. Relat.*, 6 (1987) 12.
- 9 S. Lembo, V. Sasso, C. Silipo and A. Vittoria, *Farmaco, Ed. Sci.*, 38 (1983) 750.
- 10 J. E. Garst, *J. Pharm. Sci.*, 73 (1984) 1623.
- 11 T. Braumann, *J. Chromatogr.*, 373 (1986) 191.
- 12 D. J. Minick, J. H. Frenz, M. A. Patrick and D. A. Brent, *J. Med. Chem.*, 31 (1988) 1923.
- 13 R. F. Rekker, *The Hydrophobic Fragmental Constant*, Elsevier, Amsterdam, 1977.
- 14 C. Hansch and A. Leo, *Substituent Constants for Correlation Analysis in Chemistry and Biology*, Wiley, New York, 1979.
- 15 A. J. Hopfinger and R. D. Battershell, *J. Med. Chem.*, 19 (1976) 569.
- 16 P. Broto, G. Moreau and C. Vanduycke, *Eur. J. Med. Chem.*, 19 (1984) 71.
- 17 A. K. Ghose and G. M. Crippen, *J. Chem. Inf. Comput. Sci.*, 27 (1987) 21.
- 18 J. L. Martin, J. M. Carretero, A. M. Sanz and I. J. Alonso-Silva, *Span. Pat.*, 8 801 664 (1988).
- 19 J. L. Martin, J. M. Carretero, A. M. Sanz and I. J. Alonso-Silva, *An. Quim. (C)*, in press.
- 20 J. Haavaldsen, V. Nordal and M. Kelly, *Acta Pharm. Suec.*, 20 (1983) 219.
- 21 T. Jacobsen, *Farmacoterapi*, 38 (1982) 45.
- 22 A. J. Leo, *J. Pharm. Sci.*, 76 (1987) 166.